



**International Journal of Biochemistry Research
& Review**

13(1): 1-16, 2016, Article no.IJBCRR.26949
ISSN: 2231-086X, NLM ID: 101654445

SCIECEDOMAIN international
www.sciencedomain.org



Partial Purification and Characterization of Phospholipase A₂ Inhibitor from *Echis ocellatus* Serum

F. A. Adamude^{1*}, A. J. Nok², N. Aliyu³ and E. Onyike²

¹Department of Biochemistry, Federal University Lafia, Nasarawa State, Nigeria.
²Department of Biochemistry, Ahmadu Bello University, Zaria, Kaduna State, Nigeria.
³Nuhu Bamalli Polytechnic, Zaria, Kaduna State, Nigeria.

Authors' contributions

This work was carried out in collaboration between all authors. Author FAA designed the study, wrote the protocol and author AJN supervised the work. Authors FAA and NA carried out all laboratories work and performed the statistical analysis. Author FAA managed the analyses of the study. Author NA wrote the first draft of the manuscript. Author EO managed the literature searches and edited the manuscript. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/IJBCRR/2016/26949

Editor(s):

(1) Mohamed Fawzy Ramadan Hassanien, Biochemistry Department, Zagazig University, Egypt.

Reviewers:

(1) M. I. Alam, Hamdard Institute of Medical Sciences & Research, Jamia Hamdard University, New Delhi, India.

(2) L. Chanhome, Chulalongkorn University, Thailand.

(3) Yan Yan, The Pennsylvania State University, University Park, Pennsylvania, USA.

Complete Peer review History: <http://www.sciencedomain.org/review-history/15603>

Original Research Article

Received 11th May 2016
Accepted 14th June 2016
Published 31st July 2016

ABSTRACT

The most effective and acceptable therapy for snakebite victims is the immediate administration of antivenin following envenomation which is limited by problems of hypersensitivity reactions in sensitive individuals and its inability to resolve the local effects of the venom. Phospholipase A₂ Inhibitor from *Echis ocellatus* Serum (PIES) was isolated, partially purified and characterized. The neutralizing protein from *E. ocellatus* serum inhibited the *E. ocellatus* (carpet viper) venom phospholipase A₂ (PLA₂) enzyme in a dose dependent manner. A two step purification process on sephadex G-200 column chromatography and DEAE- cellulose chromatography gave an active fraction that inhibited the venom PLA₂ by 78%. The result from SDS-PAGE showed the inhibitor to

*Corresponding author: E-mail: fatteeyjibamin@gmail.com;

be a 24.98kDa protein and its kinetic study revealed a mixed pattern of inhibition on the carpet viper venom PLA₂ with an estimated K_i values of 3.8%(v/v) to 7.3%(v/v). The study was carried out at the Department of Biochemistry, Faculty of Science, Ahmadu Bello University Zaria, Nigeria from June 2011 to August 2012. The relevance of these findings towards understanding the biochemistry of carpet viper envenomation and the development of a novel antivenin drug in future targeting the activity of PIES are discussed.

Keywords: Envenomation; antivenin; carpet viper; phospholipase A₂; neutralising protein.

1. INTRODUCTION

Snakebite is a global medical health problem especially in the rural areas of the tropics with about 40,000 deaths and 375,000 cases of permanent physical disabilities worldwide annually [1,2]. The annual snakebite incidence in northern Nigeria has been estimated to be 497 per 100,000 populations with 12.2% mainly due to the carpet viper, *Echis ocellatus* [3]. Based on hospital records, the four most deadly northern Nigerian snakes are *Echis ocellatus*, *Naja nigricollis*, *Bitis arietans* and *Naja katiensis* [4]. The incidence of snakebite is often associated with agricultural activities with the highest at the beginning of the rainy season. The other important factor responsible for the increase in snakebite cases is flood which drives out snakes from their burrows [5].

Snake venom, the most complex of all poisons is a mixture of enzymatic and non enzymatic toxic compounds as well as other nontoxic proteins, non proteins including carbohydrates and metals all stored in the poison gland. The pathophysiologic base for morbidity and mortality is the disruption of normal cellular functions by these enzymes and toxins. Some of these proteins include enzymes like phospholipase A₂, B, C, D, hydrolases, hyaluronidases, phosphatases, esterases, phosphodiesterase, neucleosidases and metalloproteases [6].

Phospholipase A₂ (PLA₂) is a lipolytic enzyme that hydrolyses the fatty acyl ester bonds at the 2-sn position of membrane phospholipids producing equimolar amounts of free fatty acid (FFA) and lysophospholipid, mainly arachidonic acid (AA); these products then become available for conversion to potent pro inflammatory mediators, such as platelet-activating factor and eicosanoids [7,8] respectively. The enzyme from snake venom is primarily used for trophic and defence functions in most species but show wide range of pharmacological activities such as neurotoxicity, myotoxicity, cardiotoxicity, with a greater impact on platelet aggregation and blood coagulation [9-11].

Venomous snakes are resistant to their own venom and several natural neutralizing proteins have been identified in their plasma [12]. In particular, PLA₂ inhibitors have been isolated from various snake sera and their primary structures have been determined [13]. However, information as regards PLA₂ inhibitor from *E. ocellatus* serum is still scanty and yet to be fully elucidated.

The main concern about the empirical use of antivenin is the limited efficacy against the local tissue damaging activities of venoms, its relative scarcity and cost factor [14,15]. Thus, specific inhibitors may alleviate the local effects of the venom, which is difficult to achieve with conventional antivenins. Therefore, in this study, we report on the purification and characterization of PLA₂ inhibitor from *E. ocellatus* serum with the view to obtain the kinetic data and establish other characteristics of the protein that would be important in achieving the said objective.

2. MATERIALS AND METHODS

2.1 Materials

Reagents were purchased from Sigma Chemical Company, St. Louis, U. S. A. Desiccator, Refrigerator, Micro pipette, Burette, Hot air oven, Water bath, SDS chamber. Twelve (12) adult *Echis ocellatus* snakes were obtained from Kaltungo, Gombe State, Nigeria and identified at the Department of Biological Sciences, Faculty of Science, Ahmadu Bello University, Zaria, Nigeria.

2.2 Collection and Preparation of Venom

Echis ocellatus identified were kept in wooden boxes. They were fed raw meat fortnightly. Venom was collected by modified milking method as described by Markfarlane. Briefly silica gel was activated at 80°C for 2 hours in an oven, placed in a dessicator and kept in a refrigerator. The snakes were restrained at the position of the joint between the last cervical vertebrae and the skull, using the thumb and the fore-finger, care

was taken to avoid strangulation. A cellophane piece was used to cover a beaker and held in position using rubber band. The second hand was used to hold the beaker and brought to make contact with the mouth of the snake, such that as the snake bites through the cellophane using its upper fangs, venom was ejected into the beaker. The venom from all the snakes were pooled and immediately placed in a desiccator with activated silica and allowed to crystallize at 4°C. The crystallized venom was then transferred into eppendorf tubes, labelled properly and kept in a deep freezer at -18°C. These were referred to as crude venom [16].

2.3 Collection and Preparation of Serum

1ml per day of blood for 3 days was obtained from each snake through the caudal vein on alternate days. The blood collection was carried out under the supervision of Dr. Ofemile Peter, Department of Veterinary Physiology, Faculty of Veterinary Medicine, Ahmadu Bello University, Zaria. After blood clotting at room temperature, the serum was separated by centrifugation at 3000xg for 5mins and kept in the refrigerator until further use [17].

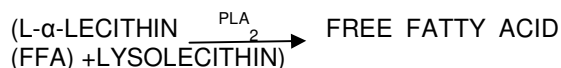
2.4 Partial Purification of Phospholipase A₂ from Crude Venom

Briefly, 2 ml of 10 mg/ml of crude *E. ocellatus* venom was loaded onto DEAE cellulose column (1.5 x 50 cm) pre-equilibrated with 50 mM phosphate buffer pH 6.8. The column was eluted stepwise with NaCl gradient (0.01 – 0.1 M) at a flow rate of 0.2 ml/min. 2 ml each of 20 fractions were collected and assayed for phospholipase A₂ activity and total protein. The PLA₂ active fractions were pooled together and loaded on sephadex G-75 column equilibrated with 50 mM phosphate buffer (pH 6.5). The column was eluted with the same buffer, maintaining a flow rate of 1ml/min. 2ml each of 20 fractions were collected and assayed for PLA₂ [18] and total protein concentration by taking absorbance at 280 nm [19].

2.5 PLA₂ Assay

Briefly, 25 µl of 1 mg/ml L-α-lecithin substrate was incubated with 10 µl of the partially purified enzyme from *E. ocellatus* venom for 10 min at 37°C. The reaction was then terminated by immersing the tube in a boiling water bath for 2 min and the amount of released free fatty acid measured titrimetrically at pH 8.0 using 20 mM

sodium hydroxide and phenolphthalein indicator. The activity of phospholipase A₂ was defined as the amount of enzyme that hydrolyses 1 µmole of fatty acids from L-α -Lecithin per minute under standard conditions.



The phospholipase A₂ activity was given as thus,

Volume of 20 mM NaOH that neutralized free fatty acid = ymL

$$0.02 \times y \times 10^{-3} = k \text{ moles}/10 \text{ min}$$

$$\text{Enzyme activity} = k \text{ moles}/10 \text{ min} = Z \text{ moles}/\text{min.}$$

2.6 Determination of Protein Concentration

The protein concentration of the partially purified PLA₂ from *Echis ocellatus* venom was determined spectrophotometrically by taking absorbance at 280 nm wavelength.

2.7 Purification of PLA₂ Inhibitor from *Echis ocellatus* Serum

PLA₂ Inhibitor from *Echis* Serum (PIES) was partially purified by a modified method. Briefly, 2 ml of serum collected from the snakes was directly fractionated at 4°C on a sephadex G-200 column, pre equilibrated with 0.05 M Tris HCl buffer, pH 8.1 containing 0.15 M NaCl. The column was eluted with the same buffer at a flow rate of 3ml/hr. 2ml of twenty fractions were collected and inhibition studies (assay for residual PLA₂ activity) was carried out and fractions exhibiting inhibitory activity against the partially purified PLA₂ from *Echis ocellatus* venom were pooled together and directly loaded on a DEAE cellulose column pre equilibrated with 0.05 M phosphate buffer, pH 6.8. The column was eluted with a stepwise linear gradient of sodium chloride (0.01- 0.1M NaCl) at a flow rate of 5min/ml. 2ml of fifty fractions collected were assayed for residual PLA₂ activity and total protein and fractions exhibiting inhibitory activity against *Echis ocellatus* PLA₂ were pooled together, dialysed and stored at -4°C until further use.

2.8 Inhibition Studies

To analyze the effect of PIES on PLA₂ activity, briefly, 10 µl of partially purified PLA₂ from *Echis ocellatus* venom, 25 µl of 1 mg/ml L-α-lecithin substrate was incubated for 10 min at 37°C with

25 μ l of various concentrations of PIES (10-50%v/v). The reaction was then terminated by immersing the tube in a boiling water bath for 2 min and the amount of released free fatty acid measured titrimetrically at pH 8.0 using 20 mM sodium hydroxide and phenolphthalein indicator. The inhibition of phospholipase A₂ activity by PIES otherwise referred to as residual PLA₂ activity was defined as the amount of residual enzyme that hydrolyzes 1 μ mole of free fatty acids from L- α -Lecithin per minute in the presence of PIES under standard conditions.

2.9 Kinetic Studies

The effect of partially purified PIES on partially purified PLA₂ activity was carried out with varying concentrations of the substrate (2-5 mg/ml) at 0%, 5%, 10% and 20% of PIES. Initial velocity values obtained were used to draw Double Reciprocal plot to determine the inhibition binding constant (K_i) and to ascertain the kind of inhibition.

2.9.1 Sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE)

Electrophoresis of the partially purified PIES under denaturing conditions was performed with 12% Acrylamide gel using a tris-glycine buffer, pH 8.3. The protein band was located by staining with Coomassie brilliant blue [20].

3. RESULTS

3.1 Purification of PLA₂ from *E. ocellatus* Venom

The results of the purification of PLA₂ from *E. ocellatus* venom are summarised on Table 1. The crude extract contained about 2.28 mg of total protein with a total activity and specific activity of 7.40 μ mol/min and 3.25 μ mol/min/mg respectively. Fractionation of the crude venom on DEAE- cellulose chromatography gave a specific activity of 21.5 μ mol/min/mg. The subsequent gel filtration on sephadex G-75 chromatography gave an active peak (Fig. 2) with a specific activity of 53.17 μ mol/min/mg, 16.36 purification fold and 43.11% recovery.

Fig. 1 shows the elution profile of *Echis* PLA₂ after ion exchange chromatography on DEAE cellulose column. An active peak eluted in fraction 13 of the void volume, had a protein concentration of 0.2 mg/ml and a corresponding enzyme activity of 4.3 μ mol/min eluted by 80 mM NaCl.

From the elution profile of *Echis* PLA₂ after gel filtration on Sephadex G-75 column (Fig. 2), an active peak (fraction 12) was found to contain about 0.06 mg/ml protein and a corresponding PLA₂ activity of 3.19 μ mol/min.

3.2 Purification of Phospholipase A₂ Inhibitor from *Echis ocellatus* Serum

The results of partial purification of phospholipase A₂ Inhibitor from *Echis ocellatus* serum (PIES) are shown in Figs. 3 and 4. From the results (Fig. 3), the elution of PIES on sephadex G-200 column chromatography produced an active peak (fraction 15) with a protein concentration of about 0.022 mg/ml and a relative inhibition of about 70% against the partially purified *E. ocellatus* PLA₂. Further purification on DEAE cellulose column (Fig. 4) gave an active peak (fraction 37) with a relative inhibition of 0.778 (77.8%) and about 0.26 mg/ml of protein.

3.3 Molecular Weight of Phospholipase A₂ Inhibitor from *Echis ocellatus* Serum (PIES)

The purity and molecular weight of the partially purified inhibitory protein were determined by Sodium Dodecyl Sulphate Polyacrylamide Gel electrophoresis (SDS-PAGE). Fig. 5 shows the electrophoretic pattern of the sample under denaturing conditions. A distinct band of the protein sample was visualised against the standard marker proteins on the gel and the molecular weight of the partially purified PIES (Band A) was estimated to be 24,986.19Da. From the plot of log of molecular weight of the marker proteins against their respective relative mobility (R_f), the molecular weight of the partially purified PIES was found to be 24,986.19Da (Fig. 6).

3.4 Characterisation of Partially Purified Phospholipase A₂ Inhibitor from *Echis ocellatus* Serum (PIES)

A study of the inhibition kinetics of partially purified PIES on partially purified PLA₂ revealed that PIES exhibited a mixed pattern of inhibition (Fig. 7) with K_M decreasing from 4.54 mg/ml without inhibitor to 3.27 mg/ml, 3.27 mg/ml and 3.12 mg/ml at 0%, 5%, 10% and 20% concentration of PIES respectively. The K_i and K_{ii} values were estimated to be 3.89% (v/v) and 7.28% (v/v) (Figs. 8 and 9), respectively.

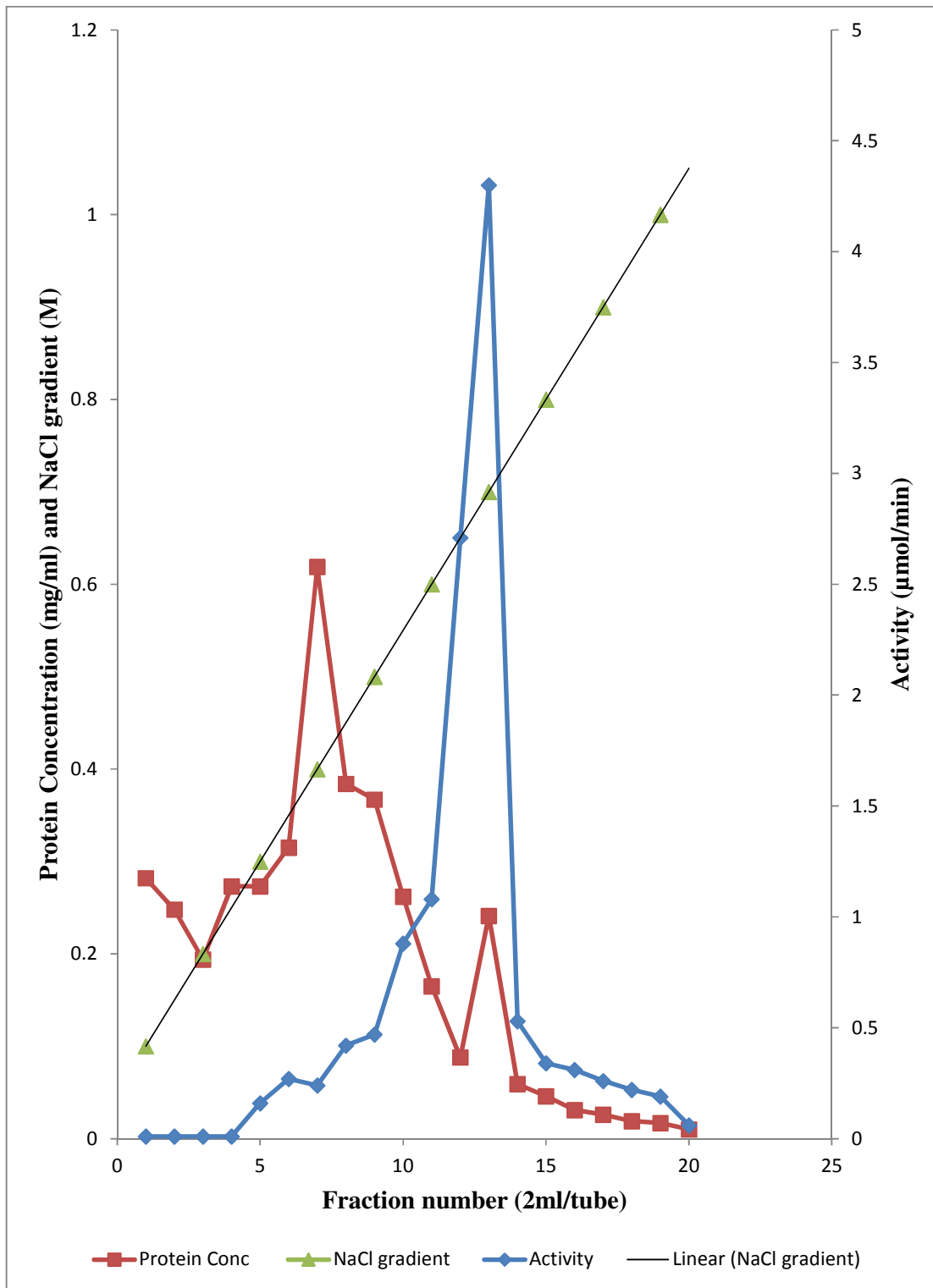


Fig. 1. Elution profile of phospholipase A₂ from *Echis ocellatus* venom on DEAE-cellulose column chromatography (1.6 x 50 cm)

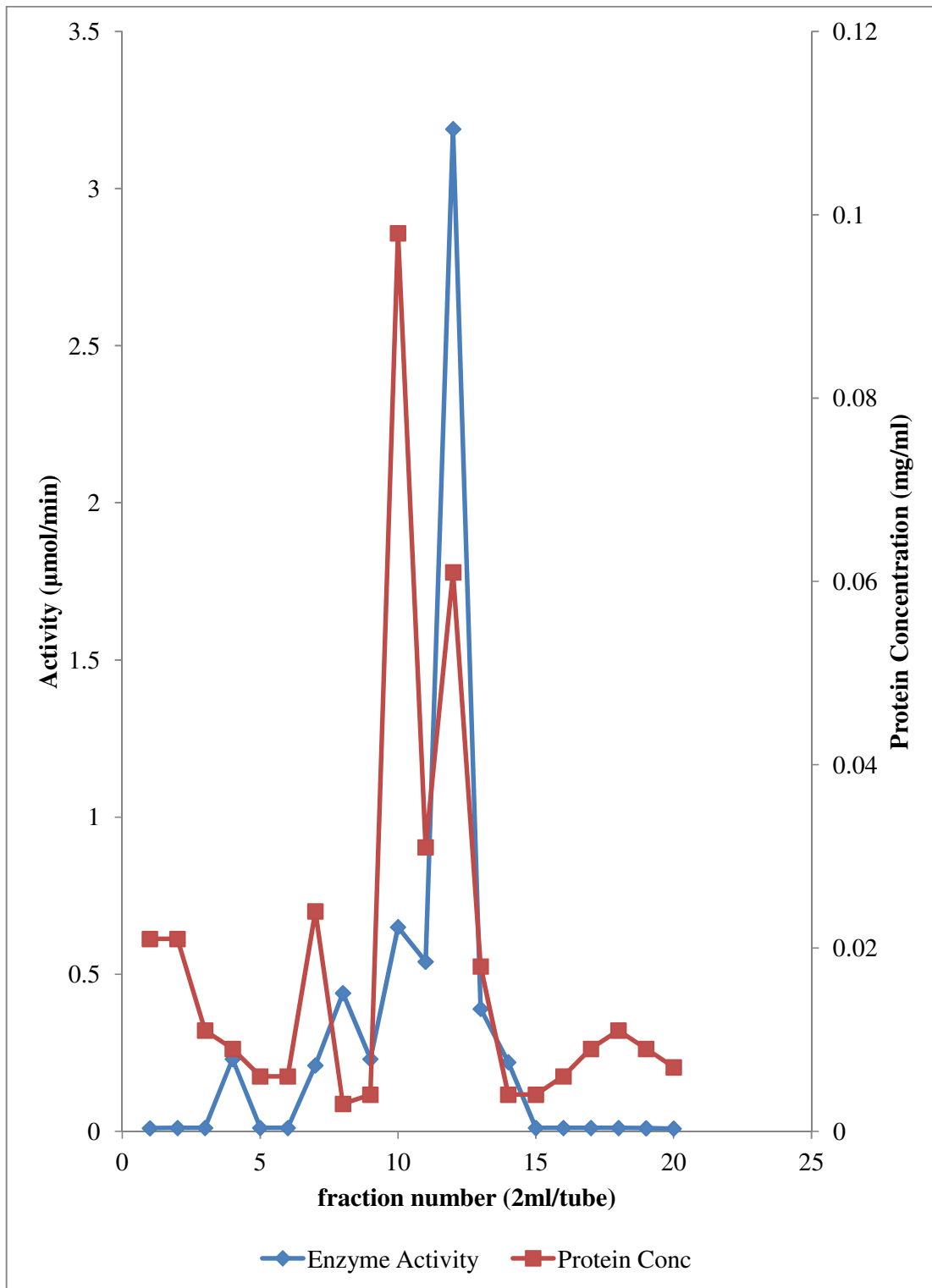


Fig. 2. Elution profile for Phospholipase A₂ from *Echis ocellatus* venom on Sephadex G-75 column chromatography (1.6 x 90 cm)

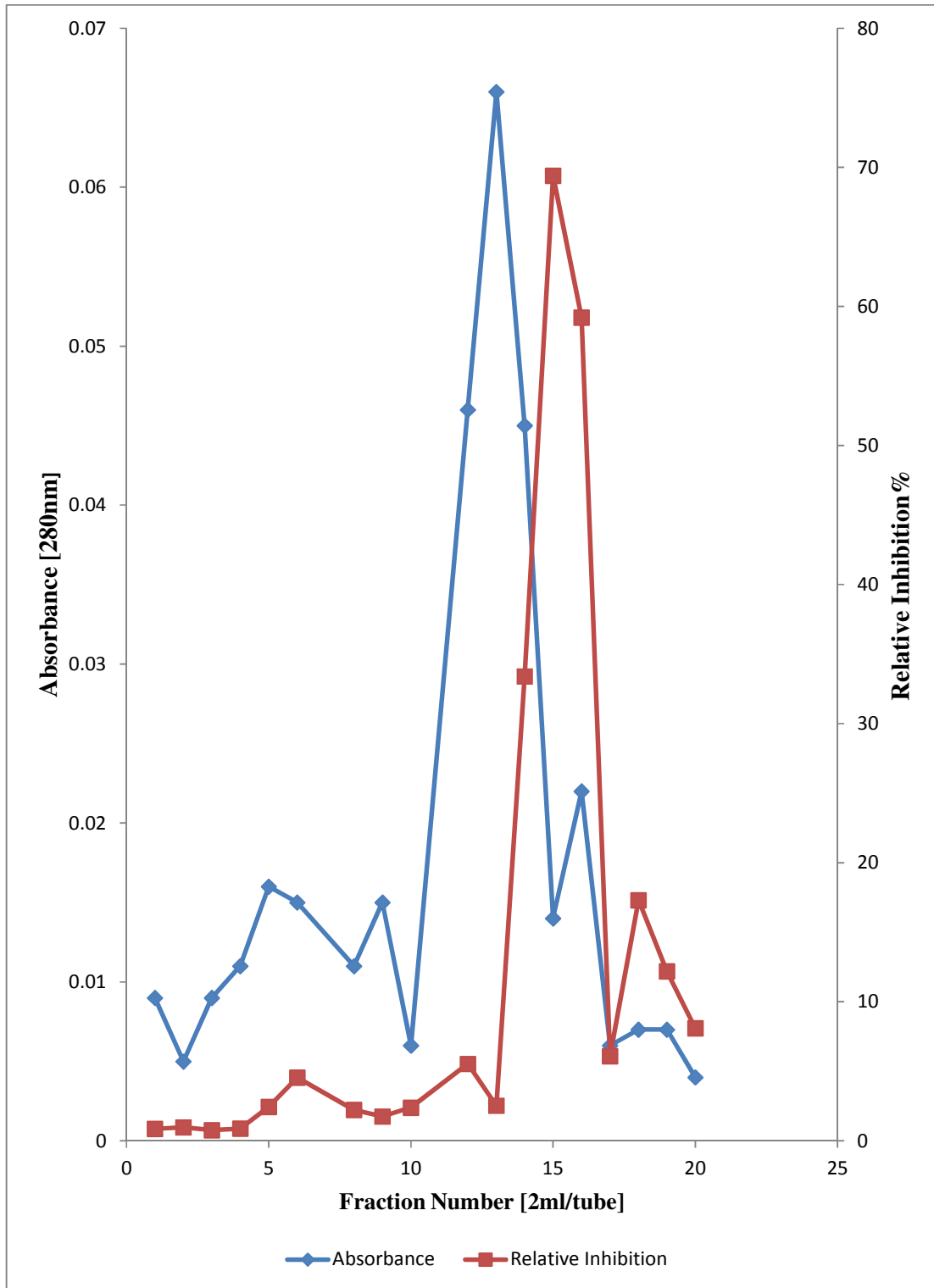


Fig. 3. Elution Profile of phospholipase A₂ inhibitor from *Echis ocellatus* serum (PIES) on Sephadex G- 200 column showing relative inhibition (%) and total protein at 280 nm

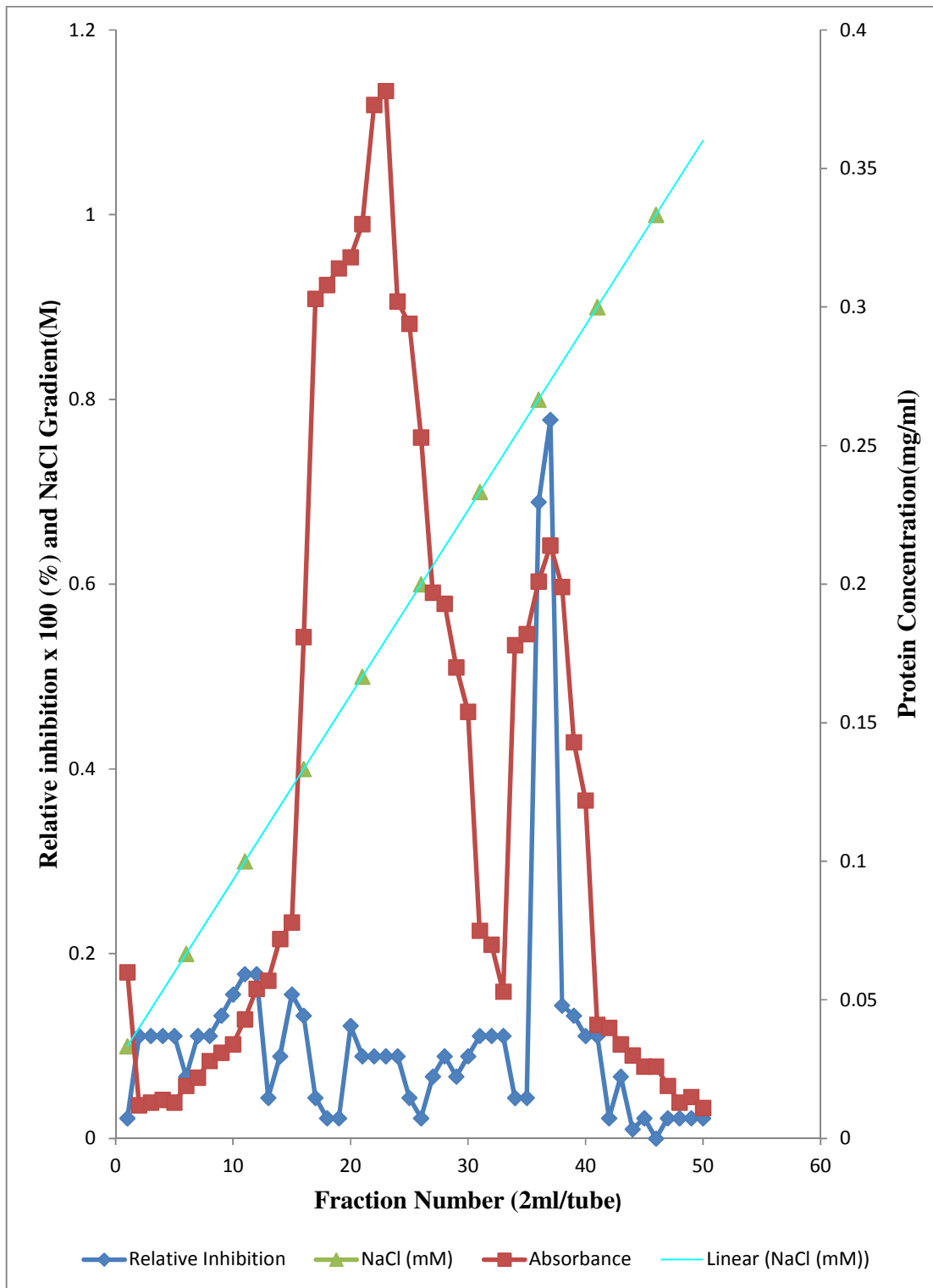


Fig. 4. Elution Profile for Phospholipase A₂ Inhibitor from *Echis ocellatus* Serum on DEAE-cellulose showing Relative Inhibition (%), total protein at 280 nm and NaCl gradient (M)

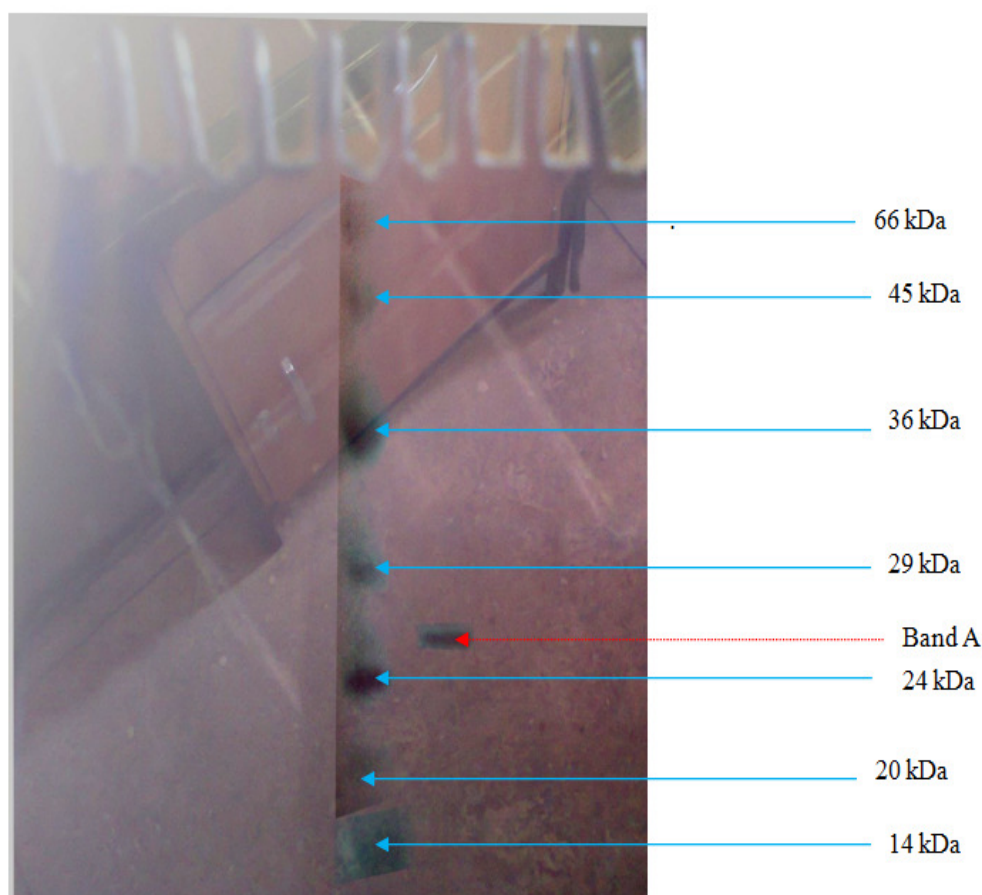


Fig. 5. Electrophoregram of Phospholipase A₂ Inhibitor from *Echis ocellatus* Serum (PIES) on Polyacrilamide Gel using Coomassie brilliant blue staining
Band A represent PIES with an estimated molecular weight of 24.98kDa

Table 1. Purification profile of phospholipase A₂ from *Echis ocellatus* venom

Purification step	Total protein (mg)	Total activity (μmol/min)	Specific activity (μmol/min/mg)	Purification fold	Percentage yield
Crude	2.28	7.40	3.25	1.00	100
Ion exchange chromatography on DEAE cellulose	0.20	4.30	21.5	6.62	58.11
Gel filtration on Sephadex G-75	0.06	3.19	53.17	16.36	43.11

*One unit of Phospholipase A₂ was defined as the amount of enzyme that hydrolyses 1 μmole of fatty acids from L-α -Lecithin per minute under standard conditions

4. DISCUSSION

Toxicological properties of snake bite are thought to be associated with enzymes especially Phospholipase A₂ (PLA₂), which is found to be its most toxic component [21]. Considering the role of PLA₂ in envenomation, understanding the characteristics of the enzyme from snake venom has raised concern for venom researchers, as it would help in the production of

effective therapeutic antivenins [22]. In this study, PLA₂ from *E. ocellatus* venom which belongs to the Class II PLA₂ predominantly found in *Viperidae* snakes was isolated and partially purified. The PLA₂ active fraction was eluted in the void volume from ion exchange chromatography on DEAE-cellulose column (Fig. 1). After gel filtration on sephadex G-75 column, an active peak eluted (Fig. 2). From the results, there was an increase in purification fold

from 6.62 to 16.36 (Table 1); while the specific activity also increased from 21.5 $\mu\text{mol}/\text{min}/\text{mg}$ to 53.17 $\mu\text{mol}/\text{min}/\text{mg}$. An increase in purification fold and specific activity of the

crude venom PLA_2 after the two purification steps could be attributed to the removal of other synergistically interacting components of the venom.

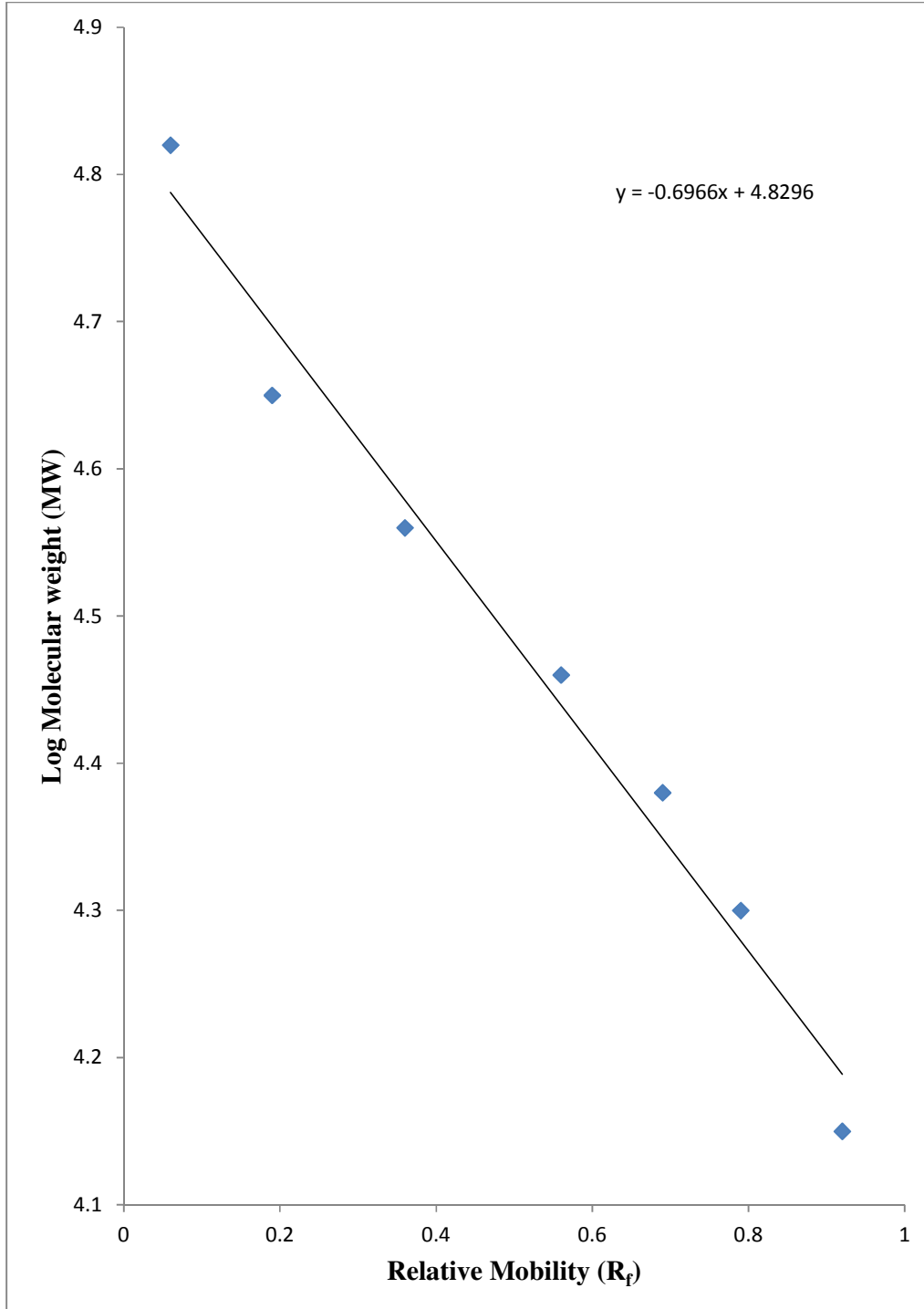


Fig. 6. A plot showing the Log of molecular weight against the R_f of the marker protein bands used to estimate the molecular weight of partially purified PIES

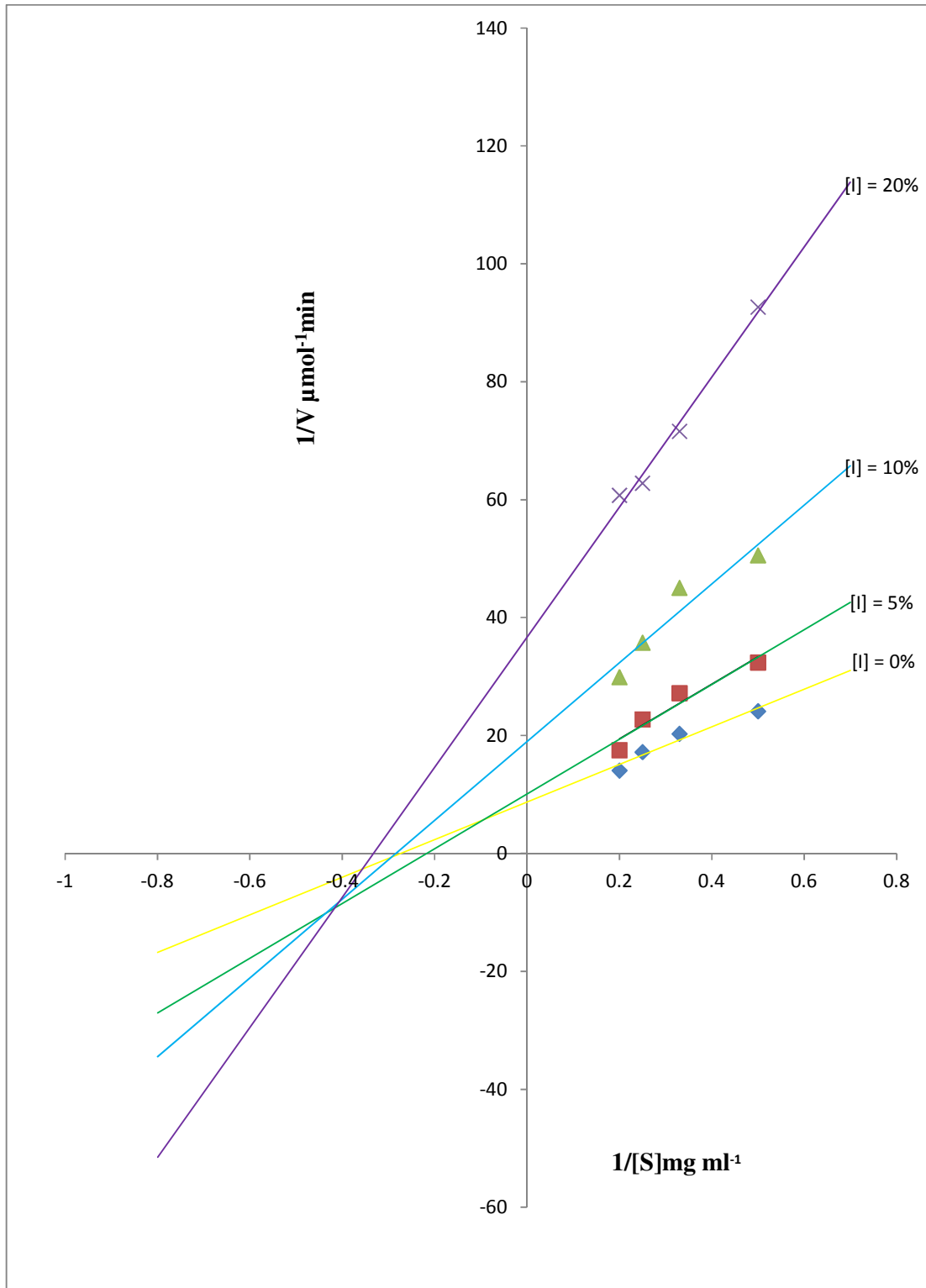


Fig. 7. Double Reciprocal plot showing the effect of partially purified PIES on partially purified *Echis* PLA₂ activity

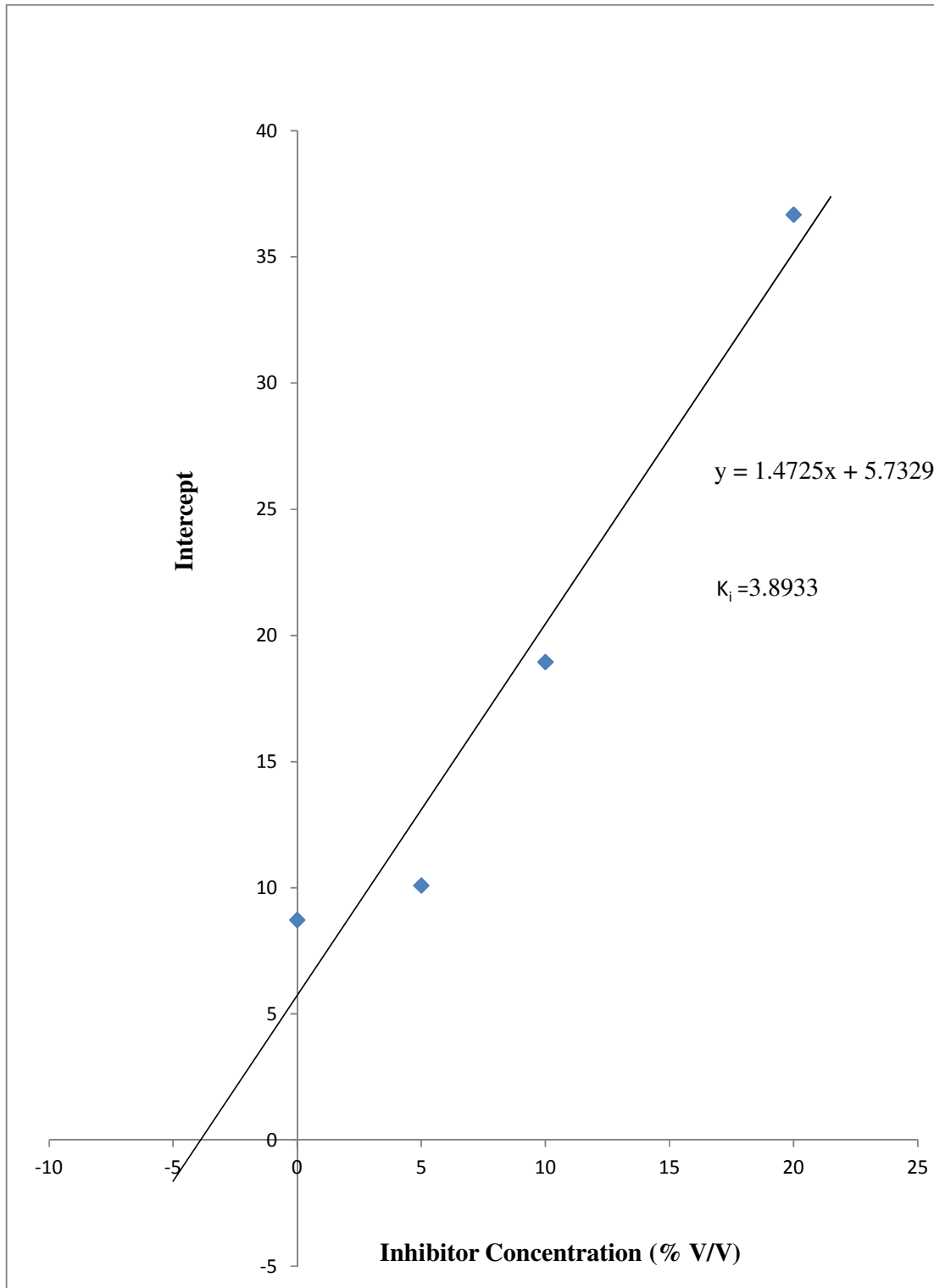


Fig. 8. Secondary plot of intercept against inhibitor concentration showing K_i

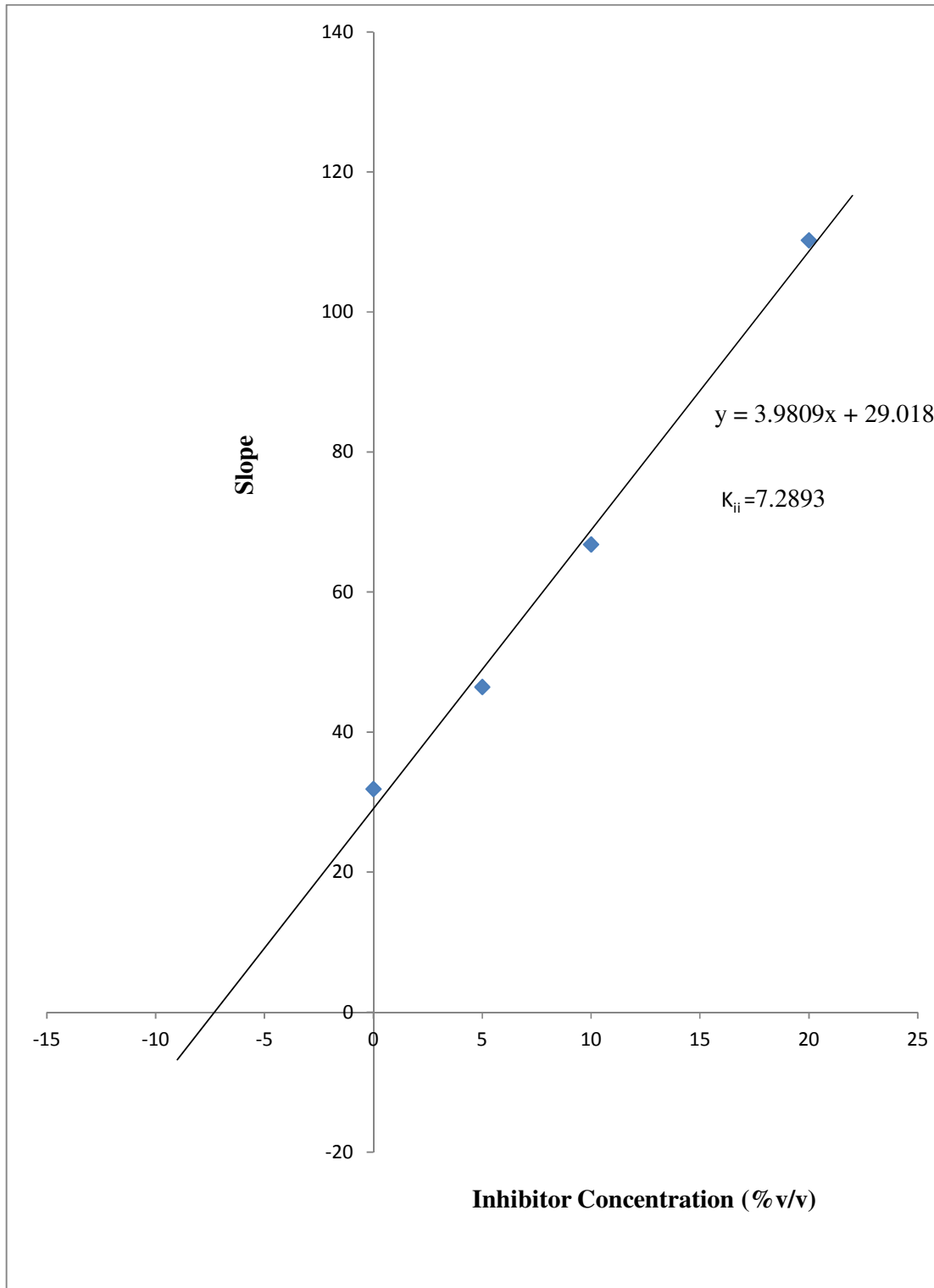


Fig. 9. Secondary plot of slope against Inhibitor concentration showing K_{ii}

Venomous snakes are resistant to their own venom and certain molecules like endogenous antitoxic serum proteins with neutralising capacity against snake venom have been previously reported [23]. In particular, PLA₂ inhibitors have been isolated from various snake sera and their primary structures have been determined [24]. In the present study, a protein PLA₂ Inhibitor from *E. ocellatus* Serum (PIES) was purified in a two-step purification process on sephadex G-200, (Fig. 3) and ion-exchange chromatography on DEAE-cellulose (Fig. 4). An active peak eluted at 80 mM NaCl inhibited the partially purified PLA₂ from the snake venom by 77.8%. The inhibitory effect of PIES may be attributed to its ability to chelate Ca²⁺, an important co-factor of PLA₂ [25]. The formation of enzyme-inhibitor or toxin-inhibitor complexes has been previously shown to be responsible for the inhibition of the PLA₂ enzyme from *T. flavoviridis* and for neutralization of the major toxins from *Naja naja atra* venom [26]. An anti-neurotoxic factor isolated from the serum of *Vipera palaestinae* forms a complex with an acidic component of the venom which participates synergistically in the neurotoxic action of this venom [27].

The purity of partially purified PIES was checked by SDS-PAGE as shown in Fig. 5. The inhibitor appeared as a single sharp band with an estimated molecular mass of about 25 kDa. This is similar to the molecular weight of PLA₂ inhibitor isolated from the serum of *Crotalus d. terrificus* snake. The isolation of a 23.6-kDa antivenin factor from *Crotalus d. terrificus* plasma has also been reported [28]. However, proteins with higher molecular weight possessing antivenin activity have also been previously reported, for example, PLA₂ inhibitors isolated from the plasma of *T. flavoviridis* [29] and *Agkistrodon b. siniticus* [30] were found to be 100 kDa and 75 kDa proteins, respectively.

Kinetic study of this research reveals that PIES exerts a mixed pattern (non competitive and uncompetitive) of inhibition on *Echis* PLA₂ (Fig. 7). This is suggestive that a site other than the active site could be involved in the inhibition since incubation of the *Echis* PLA₂ with PIES in ice for several hours did not reverse the inhibition. The result of this study is consistent with the findings that the leaves extract of *Ceiba pentandra* exerts a mixed pattern of inhibition on *Echis* PLA₂ [31]. This may also likely suggest that the PIES-PLA₂ interaction involves strong covalent forces affecting the structure [32]. The

K_i values (3.8933-7.2893%) from this study are indicative of a relatively high affinity of the inhibitor for the enzyme (Figs. 8 and 9).

5. CONCLUSIONS

From this study, it is concluded that *E. ocellatus* serum contains phospholipase A₂ (PLA₂) inhibitor with biochemical characters similar to other *Viperidea* PLA₂ inhibitors. It is also envisaged that PLA₂ inhibitor from *E. ocellatus* serum (PIES) might prove to be useful in the treatment of the various diseases in which PLA₂ enzymes have been implicated.

ACKNOWLEDGEMENTS

I wish to acknowledge the immense contributions of Mal. Shehu Shika, (Center for Energy Research and Training, Zaria, Nigeria) and Dr. Ofemile Peter of Department of veterinary Physiology, ABU Zaria, Nigeria for their skillful contributions towards the success of this work. There was no funding by any organization or agency.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Fortes-Dias CL, Fonseca BCB, Kochva E, Diniz CR. Purification and properties of an antivenom factor from the plasma of the South American rattlesnake (*Crotalus durissus terrificus*). *Toxicon*. 1991;29:997-1008.
2. Mahanta M, Mukherjee AK. Neutralization of lethality, myotoxicity and toxic enzyme of *Naja kaouthia* venom by *Mimosa pudica* root extracts. *J Ethnopharm*. 2001;75:55-60.
3. Ramar PS, Ponnampalam G, TK Chow V. Therapeutic application of natural inhibitors against snake venom phospholipase A₂. *Current Trends*. 2012;8(1). Available:www.bioinformation.net
4. EchiTAB. Annual medical records of antivenom treatment center, EchiTAB Anti-Snake Study Group UK/Nigeria, General Hospital, Kaltungo. Gombe State. Nigeria. 2008;2:1-6.
5. WHO. Guidelines for the production, control and regulation of snake. Antivenom Immunoglobulins. 2008;2:11.

6. Sallau AB, Ibrahim MA, Saliu A, Patrick FU. Characterization of phospholipase A₂ (PLA₂) from *Echis ocellatus* venom. *Afri J Biochem Res.* 2008;2(4):098-101.
7. Hasson SS, Theakston RDG, Harrison RA. Cloning of a prothrombin activator-like metalloproteinase from the West African saw-scaled viper, *Echis ocellatus*. *Toxicon.* 2003;42:629-634.
8. Ibrahim S, Nok AJ, Abubakar MS, Sarkiyayi S. Efficacy of Di-n-octyl Phthalate anti venom isolated from *Ceiba pentandra* leaves extract in neutralization of *Echis ocellatus* venom. *Appl Sci J.* 2012;4(15):2382-2387.
9. Higuchi DA, Barbosa CMV, Bincoletto C, Chagas JR, Magalhaes A, Richardson M, et al. Purification and partial characterization of two phospholipases A₂ from *Bothrops leucurus* (white tailed-jararaca) snake venom. *Biochemie.* 2007; 89:319-328.
10. Huang P, Mackessy SP. Biochemical characterization of phospholipase A₂ (trimorphin) from the venom of the *Sonoran Lyre Snake Trimorphodon biscutatus lambda* (family Colubridae). *Toxicon.* 2004;44:27-36.
11. Soares AM, Ticli FK, Marcussi S, Lourenço MV, Januário AH, Sampaio SV. Medicinal plants with inhibitory properties against snake venoms. *Current Med Chem.* 2005; 12(22):2625-2641.
12. Shashidharamurthy R, Kemparaju K. A neurotoxic phospholipase A₂ variant: Isolation and characterization from eastern regional Indian cobra (*Naja naja*). *Toxicon.* 2006;47:727-733.
13. Biondo R, Soares AM, Bertoni WB, Franca SC, Periera AM. Direct organogenesis of *Mandevilla illustris* (Vell) Woodson and effect of its aqueous extracts on the enzymatic and toxic activities of *Crotalus durissus terrificus* snake venom. *Plant Cell Rep.* 2005;22 549.
14. Sallau AB, Njoku GC, Olabisi AR, Wurocheke AU, Abdulkadir AA, Isah S, et al. Effect of *Guiera senegalensis* leaf extract on some *Echis carinatus* venom enzymes. *J Med Sci.* 2005;5(4):280-283.
15. Yang CC. Structure-function relationship of PLA₂ from snake venoms. *Journal of Toxicology-Toxin Review.* 1994;13:125-127.
16. Nobuhisa I, Inamasu S, Nakai M, Tatsui A, Mimori T, Ogawa T, et al. Characterization and evolution of the gene encoding a *Trimeresurus flavoviridis* serum protein that inhibits basic phospholipase A₂ isozymes in the snake's venom. *Eur J Biochem.* 1997;249:838-845.
17. Pugh RNH, Theakston RDG. Incidence and Mortality of snakebite in savannah Nigeria. *Lancet.* 1980;11(29):1181-1183.
18. Scott DL, Sigler PB. Structure and catalytic mechanism of secretory PLA₂. *Advances in Prot Chem.* 1994;43:53-58.
19. Bhat MK, Gowda TV. Purification and characterization of myotoxic phospholipase A₂ from Indian cobra (*Naja naja*) venom. *Biochem Int.* 1989;25:1023-1034.
20. Bharati K, Hasson SS, Oliver J, Laing GD, Theakston RDG, Harrison RA. Molecular cloning of phospholipase A₂ from venom gland of *Echis carpet* vipers. *Toxicon.* 2003;41:941-947.
21. Markfarlane RG. Russel's viper venoms. 193-1964. *Bri J Haem.* 1967;13: 437-451.
22. Shao J, Shen H, Havsteen B. Purification, characterization and binding interactions of the Chinese cobra (*Naja naja atra*) serum antitoxic protein CSAP. *Biochem J.* 1993;293:559-566.
23. Warrell DA, Davidson NMCD, Greenwood BM, Ormerod LD, Pope HM, Watkins BJ, et al. Poisoning by Bites of the Saw-Scaled or Carpet Viper (*Echis carinatus*) in Nigeria. *J Int Med.* XLVI. 1977;181:33-62.
24. Nok AJ, Esievo KAN, Ibrahim S, Ukoha AI Ikediobi ICO. Phospholipase A₂ from *Trypanosoma congolense*: characterization and haematological properties. *Cell Biochem Func.* 1993;11: 125-130.
25. Ohkura N, Inoue S, Ikeda K, Hayashi K. Isolation and amino acid sequence of a phospholipase A₂ inhibitor from the blood plasma of *Agkistrodon blomhoffii siniticus*. *J Biochem.* 1993;113: 413-419.
26. Leonard. O. Workshop on management of snakebites. *WHO Myanmar Newsletter.* 2010;2:7.
27. Perales J, Villela C, Domont GB, Choumet V, Saliou B, Moussatche AH, et al. Molecular structure and mechanism of action of the crotoxin inhibitor from *Crotalus durissus terrificus* serum. *Eur J Biochem.* 1998;227:19-26.
28. Fuly AL, Miranda AP, Zingali RB, Guimares JA. Purification and

- characterization of a phospholipase A₂ isoenzyme isolated from *Lachesis muta* snake venom. *Biochem. Pharmacol.* 2002;63:1589-1597.
29. Ovadia M, Kochva E, Moav B. Biochemical characterization and pharmacological properties of a phospholipase A₂ myotoxin inhibitor from the plasma of the snake *Bothrops asper*. *Bioch. Biophys Acta.* 1977;491:370-386.
30. Antony G, Rinku D, Sumana S, Roshnara M, Sangahamitra M, Shamik P, Aparna, G. Laboratory of toxicology and experimental pharmacothermodynamics. *Ind J Biol.* 2010;48:865-879
31. Kogaki H, Inoue S, Ikeda K, Samejima Y, Omori-Satoh T, Hamaguchi K. Isolation and fundamental properties of the phospholipase A₂ inhibitor from the blood plasma of *Trimeresurus flavoviridis*. *J Biochem.* 1989;106:966-971
32. Nok AJ, Balogun E, Lori JA, Abubakar MS. Inhibition of *Naja nigricollis* venom acidic phospholipase A₂ catalysed hydrolysis of ghost red blood cells by columbin. *J Enz Inh Med Chem.* 2002;17(1):55-59.

© 2016 Adamude et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:
<http://sciencedomain.org/review-history/15603>